

Supplementary materials for the article:

Muccee F. et al. Whole Genome Shotgun Sequencing-Based Insights into the Benzene and Xylene Degrading Potentials of Bacteria.

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Table SI

Characterization of benzene and xylene-degrading bacteria based on ribotyping, growth phases, biochemical characteristics, and organic compound removal efficiencies and GC-MS based analysis.

No.	Isolate identified (NCBI accession number)	Growth phases				Biochemical characteristics	Benzene/xylene removal efficiency	Pathways identified
		Lag	Log	Static	Death			
Benzene degrading bacteria								
1	<i>Paracoccus aestuarii</i> PUB1 (OR272055)	0–23	24–47	48–48	49 – onwards	ADH, ODC, LDC, TET, LIP, KSF, SBL, PRO, GGT, PYR, ADON	31 mg/l per 47 hour	Benzene methylation pathway, benzene degradation via benzaldehyde and via carboxylation, meta and ortho cleavage pathways, benzene degradation via phenol
2	<i>Bacillus tropicus</i> PUB2 (OR272056)	0–10	10–23	24–40	41 – onwards	ODC, LDC, LIP, KSF, SBL, GUR, PRO, GGT, PYR, ADON	30 mg/l per 23 hour	
3	<i>Bacillus albus</i> PUB3 (OR272059)	0–23	23–47	48–49	50 – onwards	ADH, ODC, LDC, TET, LIP, KSF, SBL, PRO, GGT, PYR, ADON	34 mg/l per 47 hour	
4	<i>Bacillus subtilis</i> PUB4 (OR272058)	0–23	23–47	48–48	49 – onwards	TET, LIP, KSF, SBL, PYR, ADON	32 mg/l per 47 hour	
5	<i>Bacillus thuringiensis</i> PUB5 (OR272061)	0–23	23–47	48–49	50 – onwards	ADH, ODC, LDC, LIP, PRO, GGT, PYR	30 mg/l for 47 hour	

No.	Isolate identified (NCBI accession number)	Growth phases				Biochemical characteristics	Benzene/xylene removal efficiency	Pathways identified
		Lag	Log	Static	Death			
6	<i>Bacillus cereus</i> PUB6 (OR272060)	0–23	23–47	48–49	50 – onwards	ADH, ODC, LDC, TET, LIP, KSF, SBL, PRO, GGT, PYR, ADON	31 mg/l per 47 hour	
Xylene degrading bacteria								
1	<i>Bacillus cereus</i> PUX1(OR272064)	0–11	12–48	49–50	51 – onwards	TRD, EST, URE, PRO, PYR, NO ₃	65 mg/l per 48 hour	xylene degradation via benzoate formation and anaerobic oxidation pathway
2	<i>Bacillus thuringiensis</i> PUX2 (OR272063)	0–11	12–48	49–50	51 – onwards	TRD, EST, URE, PRO, PYR, NO ₃ , GLU	40 mg/l per 48 hour	
3	<i>Bacillus cereus</i> PUX3 (OR272072)	0–23	24–48	49–50	51 – onwards	TRD, URE, PRO, PYR, NO ₃	30 mg/l per 48 hour	
4	<i>Bacillus mycoides</i> PUX4 (OR272071)	0–11	12–48	49–50	51 – onwards	TRD, URE, PRO, PYR, NO ₃	50 mg/l per 48 hour	
5	<i>Bacillus cereus</i> PUX5 (OR272073)	0–23	24–48	49–50	51 – onwards	TRD, EST, URE, PRO, PYR, NO ₃	35 mg/l per 48 hour	

ADH – Arginine dihydrolase, ADON – adonitol fermentation test, EST – triglyceride test, GGT – hydrolysis of γ -glutamyl- β -naphthylamide hydrolysis test, GLU – glucose test, GUR – β -glucuronidase test, KSF – sugar aldehyde utilization test, LDC – lysine decarboxylase test, LIP – lipase detection test, NO₃ – sodium nitrate test, ODC – ornithine decarboxylase test, PRO – proline- β -naphthylamide test, PYR – pyrrolidonyl- β -naphthylamide, SBL – sorbitol fermentation test, TET – aliphatic thiol utilization test, TRD – aliphatic diol test, URE – urea test

Table II
Raw data statistics calculated for present study the bacterial DNA consortium during whole
genome sequencing analysis

Sample ID	Total bases (bp)	Total reads	GC (%)	AT (%)	Q20 (%)	Q30 (%)
Bacterial DNA	2,232,222,732	14,782,932	54.9	45.1	93.9	86.2



Fig. S1. Quality control analysis of the DNA consortium sample by running on agarose gel electrophoresis.